

0957-4166(94)00316-5

Synthesis of α-L-Fucopyranosyl Disaccharides with Thioglycosidic Linkages and Characterization of α-L-Fucosidases from Bovine Kidney and Epididymis by their Inhibitory Activities

Hironobu Hashimoto,* Kazuya Shimada, and Shigeomi Horito[†]

Department of Life Science, Faculty of Bioscience and Biotechnology, Tokyo Institute of Technology, Nagatsuta, Midori-ku, Yokohama 227 Japan

Abstracts: Sulfur-linked disaccharide analogs of three α -L-fucopyranosyl 2-acetamido-2-deoxy- β -D-glucopyranosides and one α -L-fucopyranosyl β -D-galactopyranoside were synthesized by substitution reactions of sulfonates and ring-opening reactions of aziridine as well as oxirane derivatives using 1-thio- α -L-fucopyranose as nucleophile. Fucosidases from bovine kidney and epididymis were inhibited by all of these disaccharide analogs and their Ki values range from 0.65 to 4.9 mM.

INTRODUCTION

Oligosaccharides containing thioglycosidic linkages are well-known and easily available chemical tools for studying behaviours of carbohydrate hydrolases such as exoglycosidases 1 and endoglycosidases, 1,2 because the linkages are not affected enzymatically and the unnatural oligosaccharides behave as inhibitors. α -L-Fucopyranosyl residue is involved in many biologically important oligosaccharides, and α -L-fucosidase (EC3.2.1.15) hydrolyzes the terminal α -L-fucosyl linkage. Gamete recognition between fucose-containing glycoconjugates and α -L-fucosidase was suggested for ascidian and mammal. The most abundant linkages of L-fucose in glycoproteins and glycolipids are $\alpha(1\rightarrow 3)$, $\alpha(1\rightarrow 4)$, and $\alpha(1\rightarrow 6)$ to 2-acetamido-2-deoxy-D-glucose (D-GlcNAc) and $\alpha(1\rightarrow 2)$ to D-galactose (D-Gal). The substrate specificity of α -L-fucosidase differs with its source and has scarcely been reported for the above described α -L-fucosyl GlcNAc and Gal disaccharides except in the case of α -L-fucosidase from human serum. In order to examine the linkage specificity of α -L-fucosidase, four kinds of sulfur-linked disaccharides 1-4 were synthesized and a significant difference in their inhibitory activities was observed.

RESULTS AND DISCUSSION

General and effective O-glycosylation methods⁷ proceed by nucleophilic attack of the hydroxyl group of the acceptor sugar on the electrophilic anomeric carbon of the donor sugar. The formation of the glycosidic linkage by coupling of the donor glycosyloxy anion as nucleophile is limited ^{7b} and unpractical. On the contrary, thioglycosidic linkage has been formed effectively by the same coupling, *i.e.*, glycosylthio anion as nucleophile, due to higher nucleophilicity of the sulfur atom compared to the oxygen atom. The strategy selected for synthesis of the S-linked disaccharides 1-4 is shown in Scheme 1, where the acceptor sugars were designed considering their reactivities as electrophile.

L =leaving groups: 6-OTs, 4-OTf, NTs(2,3-aziridine), O(2,3-oxirane)

Scheme 1. Synthetic strategy of S-linked \(\alpha\)-fucopyranosyl disaccharides.

In this strategy the anomeric configuration is maintained during the coupling, because the anomerization of the glycosylthio anion is very slow. In order to utilize this benefit an efficient preparative method of 1-thio- α -L-fucopyranose triacetate (7) is necessary. Matta et al. reported⁸ conversion of L-fucose to the β -anomer of 6 via 2-S- β -L-fucosyl-2-thiopseudourea and anomerization of the β -anomer of 6 to 7, which, however, was not isolated. After unsuccessful trials of β -L-fucopyranosyl chloride preparation⁹ and addition of thioacetic acid in the presence of tert-BuOOH¹⁰ to 2-hydroxy-L-fucal triacetate, the following facile method was found. Condensation of L-fucose (5) with thioacetic acid in the presence of hydrogen chloride at room temperature for 4 h, followed by acetylation, gave 1-thio- α -L-fucopyranose tetraacetate (6) in 31% yield. The corresponding β -pyranose and α , β -furanose isomers were also formed. Selective S-deacetylation of 6 with 2-aminoethanethiol in the presence of DL-1,4-dithiothreitol gave 1-thiol 7 in 89% yield. Both 6 and 7 could be used as nucleophiles for S-linked disaccharide synthesis.

Me OH
$$a, b$$
 AcO OAC $C = 6$ R = AC 7 R = H

Scheme 2. Preparation of 1-thio-α-L-fucopyranose tetraacetate. Reagents and conditions: (a) AcSH, HCl; (b) Ac₂O, pyridine, DMAP, 31%; (c) 0.5 eq. 1,4-dithiothreitol 1.1 eq. H₂NCH₂CH₂SH / CH₃CN, 56°C, 89%.

The $\alpha(1\rightarrow 6)$ thiofucosyl linkage was formed as planned highly efficiently by the substitution reaction of a 6-O-tosyl derivative (10) of GlcNAc in DMF with the thiolate nucleophile, formed by treatment of 6 with sodium hydride, to give 11 in 99% yield. The 6-tosylate 10 was prepared from allyl 2-acetamido-2-deoxy- β -D-

glucopyranoside 8 in 35% total yield via 6-tert-butyldimethylsilyl (TBDMS) ether 9. O-Deacetylation of the disaccharide 11 gave the $\alpha(1\rightarrow 6)$ linked isomer 1 in 31% overall yield from 8.

$$R^{2}O$$
 $R^{2}O$
 R

Scheme 3. Synthesis of $\alpha(1\rightarrow 6)$ linked disaccharide.

Reagents and conditions: (a) 1) TBDMSCl, imidazole / DMF; 2) Ac₂O, DMAP / pyridine, 61% (2 steps). (b) 1) 60% AcOH, 70°C; 2) TsCl / pyridine, 60% (2 steps). (c) 1)7, NaH / DMF; 2) Ac₂O / pyridine, DMAP, 99%; 3) NaOMe / MeOH, 91%.

For the construction of $\alpha(1\rightarrow 4)$ linkage allyl 2-azido-2-deoxy- β -D-galactopyranoside (12) was selected as a precursor of GlcNAc moiety. Its 4-triflate 14 was prepared *via* the corresponding 3,6-bis-O-TBDMS derivative 13, which was deduced to be more reactive than 3,6-di-O-pivaloyl analog, which was previously¹¹ used for preparation of a 4-thio derivative of GlcNAc. The same nucleophilic substitution of 14 with 7 gave the S-linked disaccharide 15 in 68% yield including the 4-O-triflylation step. O-Desilylation of 15, followed reduction of the azido group with hydrogen sulfide, purification as the peracetate 17 and O-deacetylation, gave the $\alpha(1\rightarrow 4)$ linked isomer 2 in 57% total yield.

Scheme 4. Synthesis of α(1-4) linked disaccharide.

Reagents and conditions: (a) TBDMSCI, imidazole / DMF, 60°C, 58%. (b) Tf₂O, pyridine, DMAP / CH₂Cl₂. (c) 1) 7,NaH/DMF; 2) Ac₂O / pyridine, DMAP, 68% (3 steps). (d) 1)Bu₄NF / THF; 2) Ac₂O, DMAP / pyridine, 91% (2 steps). (d) 1)H₂S, pyridine, / H₂O; 2) Ac₂O, DMAP / pyridine, 97% (2 steps); 3) NaOMe / MeOH, 65%.

It is rather well known¹² that nucleophilic substitutions at C-3 of GlcNAc derivatives give often cumbersome results. On the other hand, an introduction of benzylthio group at C-3 position via 2,3-epimino derivative was reported to be highly regionselective. ¹² 4,6-O-Benzylidene-2,3-epimino-β-D-allopyranoside (19)

was prepared in 52% yield (2 steps) via the 3-mesylate 18 in the same manner as reported by Meyer zu Reckendorf and Lenzen¹³ for the corresponding methyl glycoside. In order to elevate the reactivity of the aziridine ring tosyl group was introduced at the nitrogen (20) and O-debenzylidenated to give a coupling partner 21 quantitatively.

The ring opening reaction of 21 with 6 in the presence of sodium methoxide in methanol at reflux, followed by acetylation, gave a mixture of the desired disaccharide 22 and its D-altro isomer 23 with a ratio of 2: 1 in 92% yield, which were separated on a column of silica gel with toluene-ethyl acetate. N-Detosylation under several reported methods, ¹⁴ where O-allyl group can survive, was unsuccessful. Then, after conversion of the aglycon moiety from allyl to propyl by catalytic hydrogenation, thus obtained $\alpha(1\rightarrow 3)$ linked disaccharide 24 was subjected to Birch reduction, followed by acetylation for purification and O-deacetylation, to obtain 3 in 81% total yield.

Scheme 5. Synthesis of $\alpha(1\rightarrow 3)$ linked disaccharide. Reagents and conditions: (a) MsCl/pyridine, 71%. (b) 1) i-PrONa/i-PrOH-dioxane, reflux, 73%; 2) TsCl/pyridine, 98%. (c) H⁺resin/80% MeOH, 60°C, quant. (d) 1) 7, NaOMe/MeOH, reflux; 2) Ac₂O/pyridine, DMAP, 22: 63%, 23: 29% (3 steps). (e) 1) Pd/C, H₂/MeOH, quant.; 2) NaOMe/MeOH, 90%. (f) 1) Na/NH₃, THF, -78°C; 2) Ac₂O, DMAP/pyridine, 85%. (g) NaOMe/MeOH, 94%.

2-Thio-D-galatopyranoside derivative was prepared in a moderate yield by mercaptonitration ¹⁵ of D-galactal triacetate with potassium thiobenzoate in the presence of ceric ammonium nitrate. Although introduction of sulfur atom at C-2 by substitution of triflate or ring-opening reaction of epoxide was reported, no examples of D-talo isomer which gives D-galactose derivative was found. Thus the latter possibility was examined using 6 as nucleophile. The coupling of 1,6:2,3-dianhydro-D-talopyranose (27) with 6 in highly concentrated solution (2.6 M for 6) in methanol in the presence of sodium methoxide gave a desired C₂-thiofucosylated D-galacto isomer 28 and its C₃-linked D-allo isomer 29 in 87% and 7% yields, respectively. The ring-opening reaction in a diluted solution (0.9 M) resulted in a prolonged reaction time and a reduced yield of 28 due to the

anomerization of 1-thio-L-fucose. Here the ring-opening reaction seems to proceed under the control of Fürst-Plattner rule. However, it is noteworthy that a distinct reverse regioselectivity in the ring-opening reaction of the corresponding D-manno isomer (C-4 epimer) of 27 with benzyl mercaptan was reported. Considering the anomeric configuration of D-galactose in natural glycoconjugates the $\alpha(1\rightarrow 2)$ linked disaccharide 28 was converted to the β -glycoside 32 via 1-acetate (30) and 1-O-trichloroacetimidate (31). The glycosidation reaction with allyl alcohol in the presence of boron trifluoride etherate gave the β -glycoside 32 predominately in 90% yield. O-Deacetylation of 32 gave the disaccharide 4 quantitatively. Here described synthetic methods, especially by the ring-opening reactions of epimino and oxirane derivatives, proved to be effective for the construction of thioglycosidic oligosaccharides.

Scheme 6. Synthesis of $\alpha(1\rightarrow 2)$ linked disaccharide. Reagents and conditions: (a) 1) 6, NaOMe / MeOH, reflux; 2) Ac₂O / pyridine, DMAP, 28: 87%, 29: 7%. (b) Ac₂O / AcOH, H₂SO₄, quant. (α : β =88:12). (c) 1) H₂NNH₂·AcOH, DMF, 50°C, 2) CCl₃CN, DBU / CH₂Cl₂ (2 steps, 78%). (d) AllOH, MS4A, BF₃·OEt₂ / CH₂Cl₂, -45°C, 98%. (e) NaOMe / MeOH, quant.

The structures of the above described disaccharide derivatives were confirmed by 1 H- and 13 C-NMR data. The position where the 1-thio- α -L-fucopyranosyl residue is introduced is clearly indicated by the higher chemical shift (from 2.8 to 3.4 ppm depending on the position and orientation). The coupling constants of the ring protons, i.e. $J_{2,3} = 10.3$ and $J_{3,4} = 10.1$ Hz for 22, $J_{2,3} = 5.6$ and $J_{3,4} = 4.3$ Hz for 23, $J_{3,4} = 5.0$ Hz for 28 and $J_{3,4} = 10.1$ Hz for 29, also support the configuration especially for the ring-opening products.

 α -L-Fucosidase are important enzymes in the metabolism of L-fucose-containing biological substances and have been investigated extensively. ¹⁸ Human serum α -L-fucosidase⁶ is one of the well-characterized glycosidase and was demonstrated to hydrolyze the $\alpha(1\rightarrow 2)$ linkage effectively. On the other hand, commercially available α -L-fucosidases from bovine kidney and epididymis ¹⁹ are known to be able to hydrolyze all the four kinds of α -L-fucopyranosyl linkages as well as p-nitrophenyl α -L-fucopyranoside, but enzymological kinetic data have not been reported. Thus our synthesized S-linked oligosaccharides were examined at first toward these α -L-fucosidases. Inhibitory activities of the S-linked oligosaccharides 1-4 were examined using p-nitrophenyl α -L-fucopyranoside as the substrate, and expressed in the term of Ki values, which were estimated based on the Lineweaver-Burk plot as summarized in Table 1. The most remarkable inhibition was observed for the $\alpha(1\rightarrow 3)$ linked disaccharide 3 with a mode of competitive inhibition. Other S-linked disaccharides 1, 2 and 4 showed mixed inhibition. As preliminary reported ²⁰ 5-thio- α -L-fucopyranosyl

analog of 4 shows a potent competitive inhibition (Ki = 30 μ M) against α -L-fucosidase from bovine epididymis, the observed mixed inhibition of 4 is deduced to be attributed to the effect of its rather high concentration (Ki = 1.1 mM). Here observed difference in the inhibitory activities of four S-linked α -L-fucopyranosyl disaccharides indicates a usefulness of these disaccharide analogs for characterization of exoglycosidases.

| Table 1. | Inhibitory | activities of | sulfur-linked | disaccharide | analogs 1 | i – 4 towa | rd α-L-fucosidases |
|----------|------------|---------------|---------------|--------------|-----------|------------|--------------------|
|----------|------------|---------------|---------------|--------------|-----------|------------|--------------------|

| Disaccharide | Ki values (mM) and inhibition mode ^a | | | | |
|--------------|---|------------------------------|--|--|--|
| analog | bovine kidney | bovine epididymis | | | |
| 1 | 4.5 (mixed, $\alpha = 9$) | 4.7 (mixed, $\alpha = 7$) | | | |
| 2 | 4.9 (mixed, $\alpha = 2$) | 3.5 (mixed, $\alpha = 4$) | | | |
| 3 | 0.65 (competitive) | 0.66 (competitive) | | | |
| 4 | $0.88 \text{ (mixed,}^b \alpha = 0.5)$ | 1.1 (mixed, $\alpha = 0.5$) | | | |

^a The inhibition modes were indicated by Lineweaver-Burk plots and replots of the slope as well as intercept versus the inhibitor concentration.

EXPERIMENTAL

General method. Melting points were determined with a Mel-Temp. apparatus and are not corrected. Optical rotations were measured using a 0.5 dm tube and a JASCO DIP-4 Polarimeter. NMR spectra were recorded at 270 MHz with a JEOL JNM-EX-270 in CDCl₃ with tetramethylsilane as a internal standard, unless stated otherwise. Column chromatography was performed on Wakogel C-300 (Wako Pure Chemical Industries, Ltd.)

2,3,4-Tri-O-acetyl-1-S-acetyl-1-thio- α -L-fucopyranose (6). Into thioacetic acid (100 mL) was introduced hydrogen chloride at 0°C. To the saturated solution was added 5 (9.84 g, 60 mmol) with stirring at 0°C and this temperature was maintained for 10 min. Then the mixture was kept at room temperature for 4 h. After evaporation in vacuo, the resulting syrup was acetylated with Ac₂O (50 mL), pyridine (50 mL), and DMAP (catalytic amount) to give 6 (6.56 g, 31%) after workup and chromatography with 10: 1 hexane-EtOAc. Compound 6 had mp 119-120°C; $[\alpha]_D^{20}$ -156 (c 0.52, CHCl₃); ¹H NMR data: δ 6.23 (d, 1 H, $J_{1,2}$ = 5.3 Hz, H-1), 5.48 (dd, 1 H, $J_{2,3}$ = 10.9 Hz, H-2), 5.29 (dd, 1 H, $J_{3,4}$ = 3.2 Hz, $J_{4,5}$ = 1.3 Hz, H-4), 5.03 (dd, 1 H, H-3), 4.06 (q, 1 H, $J_{5,6}$ = 6.4 Hz, H-5), 2.41 (s, 3 H, SAc), 2.18, 2.01, 2.00 (3 s, 9 H, 3 OAc); ¹³C NMR data: δ 192.2, 170.5, 169.9, 169.6, 81.2, 70.4, 69.3, 69.1, 66.3, 31.5, 20.7, 20.6, 16.1; Anal. Calcd for $C_{14}H_{20}O_8S$: C, 48.27; H, 5.79. Found: C, 48.12; H, 5.90.

2,3,4-Tri-O-acetyl-1-thio- α -L-fucopyranose (7). To a solution 6 (220 mg, 0.632 mmol) and DL-dithiotheritol (44 mg, 0.29 mmol) in acetonitrile (5 mL) was added cysteamine (52 mg, 0.67 mmol) and the solution was kept for 25 min at 55°C. Chloroform was added and the mixture was washed with ice water, dried (MgSO₄) and evaporated in vacuo. The residue was chromatographed with 1:2 hexane-EtOAc to give 7 (172 mg, 89%). Compound 7 had $[\alpha]_D^{20}$ -162 (c 1.00, CHCl₃); ¹H NMR data: δ 5.99-5.96 (m, 1 H, H-1), 5.31-5.21 (m, 3 H, H-2, 3, 4), 4.53 (q, 1 H, $J_{5,6}$ = 6.6 Hz, H-5), 2.17, 2.09, 2.01 (3 s, 9 H, 3 Ac), 1.80 (d, 1 H, $J_{5,6}$ = 5.3 Hz, SH), 1.16 (d, 1 H, H-6); ¹³C NMR data: δ 170.6, 170.1, 77.7, 70.9, 68.1, 67.6, 65.5, 20.9,

^b Linear mixed-type inhibition²¹ which may be considered as a mixture of partial competitive inhibition and pure noncompetitive inhibition.

20.8, 20.7, 16.0; Anal. Calcd for C₁₂H₁₈O₇S: C, 47.05; H, 5.92. Found: C, 47.32; H, 6.03.

Allyl 2-acetamido-3,4-di-O-acetyl-6-O-tert-butyldimethylsilyl-2-deoxy- α -D-glucopyranoside (9). To a mixture of 8^{22} (3.66 g, 14.0 mmol) and imidazole (2.48 g, 36.4 mmol) in DMF (11 mL) was added a solution of tert-butylchlorodimethylsilane (2.74 g, 18.2 mmol) in DMF (5 mL) at 0°C and the solution was kept at this temperature for 30 min. The mixture was treated with Ac₂O (10 mL), pyridine (15 mL), and DMAP to give 9 (3.93 g, 61%) after workup. Compound 9 had mp 212-213°C; $[\alpha]_D^{20}$ -3.2 (c 1.34, CHCl₃); ¹H NMR data: δ 5.93-5.79 (m, 1 H, OCH₂CH=CH₂), 5.62 (d, 1 H, $J_{2,NH}$ = 8.6 Hz, NH), 5.30-5.17 (m, 3 H, H-3, OCH₂CH=CH₂), 4.99 (t, 1 H, $J_{3,4}$ = 9.2 Hz, $J_{4,5}$ = 9.9 Hz, H-4), 4.63 (d, 1 H, $J_{1,2}$ = 8.6 Hz, H-1), 4.37-4.30 (m, 1 H, OCH₂CH=CH₂), 4.12-4.05 (m, 1 H, OCH₂CH=CH₂), 3.91(dt, 1 H, $J_{2,3}$ = 10.7 Hz, H-2), 3.70 (d, 2 H, $J_{5,6}$ = 4.2 Hz, H-6), 3.53 (dt, 1H, H-5), 2.03, 2.02, 1.95 (3 s, 9 H, 3 Ac), 0.89 (s, 9 H, t-Bu), 0.06, 0.05 (2 s, 6 H, 2 Me); ¹³C NMR data: δ 171.1, 170.1, 169.3, 133.7, 117.6, 99.5, 74.7, 72.9, 69.4, 69.2, 62.6, 54.5, 25.8, 23.3, 20.7, 18.3, -5.4; Anal. Calcd for C₂₁H₃₇NO₈Si: C, 54.88; H, 8.11; N, 3.05. Found: C, 54.60; H, 8.28; N, 2.97.

Allyl 2-acetamido-3,4-di-O-acetyl-2-deoxy-6-O-p-toluenesulfonyl- β -D-glucopyranoside (10). To 70% acetic acid (15 mL) was added 9 (1.40 g, 3.05 mmol), the mixture was kept for 1.5 h at 70°C and evaporated in vacuo. To a solution of the residue in pyridine (5 mL) was added p-toluenesulfonyl chloride (0.70 g, 3.7 mmol) with stirring. The mixture was kept at room temperature for 3 h and poured into methanol. The precipitate was filtered and recrystallized from EtOH-hexane to give 10 (0.91 g, 60%). Compound 10 had mp 177-179°C (decomp.); $[\alpha]_D^{24}$ -1.8 (c 1.30, CHCl₃); 1 H NMR data: δ 7.76, 7.36 (each d, each 2 H, J = 8.3 Hz, aromatic in Ts), 5.89-5.75 (m, 1 H, OCH₂CH=CH₂), 5.60 (d, 1 H, J_{2,NH} = 8.9 Hz, NH), 5.30-5.17 (m, 3 H, OCH₂CH=CH₂, H-3), 4.90 (t, 1 H, J_{3,4} = J_{4,5} = 9.6 Hz, H-4), 4.65 (d, 1 H, J_{1,2} = 8.5 Hz, H-1), 4.29-4.22 (m, 1 H, OCH₂CH=CH₂), 4.15-3.98 (m, 3 H, H-6a, 6b, OCH₂CH=CH₂), 3.82 (dt, 1 H, J_{2,3} = 10.6 Hz, H-2), 3.79-3.72 (m, 1 H, H-5), 2.46 (s, 3 H, Me in Ts), 2.02, 2.00, 1.94 (3 s, 9 H, 3 Ac); 13 C NMR data: δ 170.8, 170.2, 169.5, 145.1, 133.3, 132.4, 129.9, 128.0, 117.8, 99.4, 72.0, 71.6, 69.8, 68.9, 68.1, 54.5, 23.3, 21.6, 20.6, 20.5; Anal. Calcd for C₂₂H₂₉NO₁₀S: C, 52.90; H, 5.85; N, 2.80. Found: C, 52.82; H, 5.74; N, 2.59.

Allyl 2-acetamido-3,4-di-O-acetyl-6-S-(2,3,4-tri-O-acetyl-α-L-fucopyranosyl)-2-deoxy-6-thio-β-D-glucopyranoside (11). To a solution of 7 (408 mg, 1.33 mmol) in DMF (1 mL) was added sodium hydride (55%, 65 mg, 1.5 mmol) at 0°C. After 2 min, 10 (443 mg, 0.887 mmol) was added to the mixture and kept for 1 h at room temperature. The mixture was treated with Ac₂O (1 mL), pyridine (1 mL), and DMAP to give 11 (55 mg, 99%) after workup and chromatography in 1 : 4 hexane-EtOAc. Compound 11 had $[\alpha]_D^{24}$ -127 (c 1.50, CHCl₃); ¹H NMR data: δ 6.03 (d, 1 H, $J_{2,NH}$ = 8.5 H, NH), 5.96-5.81 (m, 1 H, OCH₂CH=CH₂), 5.74 (d, 1 H, $J_{1,2}$ = 5.3 Hz, H-1), 5.34-5.19 (m, 6 H, H-3, H-2',3',4', OCH₂CH=CH₂), 4.92 (t, 1 H, $J_{3,4}$ = $J_{4,5}$ = 9.4 Hz, H-4), 4.72 (d, 1 H, $J_{1,2}$ = 8.5 Hz, H-1), 4.43 (q, $J_{5',6'}$ = 6.6 Hz, H-5'), 4.38-4.31 (m, 1 H, OCH₂CH=CH₂), 4.11-4.04 (m, 1 H, OCH₂CH=CH₂), 3.86 (dt, 1 H, $J_{2,3}$ = 10.6 Hz, H-2), 3.62(dt, 1 H, $J_{5,6a}$ = 8.4 Hz, $J_{5,6b}$ = 3.0 Hz, H-5), 2.76 (dd, 1 H, $J_{6a,6b}$ = 13.9 Hz, H-6a), 2.64 (dd, 1 H, H-6b), 2.17, 2.07, 2.04, 2.03, 1.99, 1.94 (6 s, 18 H, 6 Ac), 1.17 (d, 3 H, H-6'); ¹³C NMR data: δ 170.6, 170.4, 170.2, 169.9, 169.8, 169.5, 133.5, 117.4, 99.4, 82.8, 74.5, 72.1, 71.9, 70.6, 69.6, 68.3, 67.7, 64.6, 54.6, 30.8, 23.1, 20.7, 20.6, 20.5, 20.4, 15.8; Anal. Calcd for C₂₇H₃₉NO₁₄S: C, 51.18; H, 6.20; N, 2.21. Found: C, 51.01; H,6.14; N, 2.25.

Allyl 2-acetamido-2-deoxy-6-S-(α-L-fucopyranosyl)-6-thio-β-D-glucopyranoside (1). A solution of 11 (452 mg, 0.713 mmol) and a 1.0 M solution of MeONa in MeOH (0.3 mL) were mixed, and stirred for 20 min at room temperature. The mixture was neutralized with Dowex 50 W-X2 (H⁺) resin and filtered. The filtrate was concentrated in vacuo to give 1 (274 mg, 91%). Compound 1 had mp 169-171°C; $[\alpha]_D^{24}$ -132 (c 0.75, H₂O); ¹H NMR (D₂O): δ 5.94-5.92 (m, 1 H, OCH₂CH=CH₂), 5.52 (d, 1 H, $I_{1',2'}$ = 5.6 Hz, H-1'), 5.36-5.26 (m, 2 H, OCH₂CH=CH₂), 4.57 (d, 1 H, $I_{1,2}$ = 8.6 Hz, H-1), 4.39-4.31 (m, 2 H, H-5, OCH₂CH=CH₂), 4.22-4.15 (m, 1 H, OCH₂CH=CH₂), 4.08 (dd, 1 H, $I_{2',3'}$ = 10.2 Hz, H-2'), 3.83 (d, 1 H, $I_{4',5'}$ = 3.0 Hz, H-4'), 3.77-3.69 (m, 2 H, H-2, 3'), 3.56-3.35 (m, 3 H, H-3, 4, 5), 3.12 (d, 1 H, $I_{6a,6b}$ = 14.2 Hz, H-6b), 2.83 (dd, 1 H, $I_{5,6b}$ = 8.9 Hz, H-6b), 2.04 (s, 3 H, Ac), 1.25 (d, 3 H, H-6'), ¹³C NMR (D₂O): δ 174.9, 133.7, 118.6, 100.5, 87.5, 76.4, 74.0, 73.5, 72.0, 70.9, 70.6, 68.1, 67.6, 56.0, 32.3, 22.5, 45.8; Anal. Calcd for C₁₇H₂₉NO₉S: C, 48.22; H, 6.90; N, 3.31. Found: C, 48.00; H, 7.09; N, 3.06.

Allyl 2-azido-3,6-di-O-tert-butyldimethylsilyl-2-deoxy-β-D-galactopyranoside (13). To a mixture of 12^{22} (584 mg, 2.38 mmol) and imidazole (0.65 g, 9.5 mmol) in DMF (2 mL) was added a solution of tert-butylchlorodimethylsilane (0.90 g, 6.0 mmol) in DMF (2 mL). The mixture was stirred for 20 h at 65°C, and diluted with benzene. The organic layer was washed with brine, dried (MgSO₄) and evaporated in vacuo. The residue was chromatographed with 20: 1 hexane-EtOAc to give 13 (655 mg, 58%). Compound 12 had $[\alpha]_D^{24}$ +4.0 (c 1.91, CHCl₃); ¹H NMR data: δ 6.01-5.86 (m, 1 H, OCH₂CH=CH₂), 5.36-5.18 (m, 2H, OCH₂CH=CH₂), 4.42-4.36 (m, 1 H, OCH₂CH=CH₂), 4.26 (d, 1 H, $J_{1,2}$ = 7.6 Hz, H-1), 4.17-4.09 (m, 1 H, OCH₂CH=CH₂), 3.91 (dd, 1 H, $J_{5,6a}$ = 6.8 Hz, $J_{6a,6b}$ = 10.1 Hz, H-6a), 3.82 (dd, 1 H, $J_{5,6b}$ = 5.9 Hz, H-6b), 3.79-3.77 (m, 1 H, H-4), 3.54-3.39 (m, 3 H, H-2, 3, 5), 2.48 (s, 1 H, OH), 0.93, 0.89 (2 s, 18 H, 2 t-Bu), 0.18, 0.13, 0.08 (3 s, 12 H, 4 Me); ¹³C NMR data: δ 133.6, 117.5, 100.8, 74.5, 73.6, 69.8, 68.2, 64.6, 61.8, 25.8, 25.7, 18.2, 18.0, -4.7, -4.9, -5.4, -5.5; Anal. Calcd for C₂₁H₄₃N₃O₅Si₂: C, 53.24; H, 9.15; N, 8.87. Found: C, 52.99; H, 8.68; N, 8.54.

Allyl 2-azido-3,6-di-0-tert-butyldimethylsilyl-4-S-(2,3,4-tri-0-acetyl- α -L-fucopyranosyl)-2deoxy-4-thio-β-D-glucopyranoside (15). To a solution of pyridine (0.7 mL, 8.7 mmol) and DMAP (18 mg, 0.15 mmol) in CH₂Cl₂ (3 mL), was added a solution of trifluoromethanesulfonic anhydride (0.50 mL, 3.0 mmol) in CH₂Cl₂ (2 mL) at 0°C, and after stirred for 30 min at this temperature then 13 (700 mg, 1.48 mmol). The mixture was stirred for 1 h at room temperature, and then diluted with CH₂Cl₂. The organic layer was washed with M hydrochloric acid, aq sodium hydrogencarbonate (saturated), dried (MgSO₄), and evaporated in vacuo to give a syrup of triflate 14. To a solution of 7 (473 mg, 1.43 mmol) in DMF (0.5 mL) was added sodium hydride (55%, 68 mg, 1.6 mmol) at 0°C under argon. After 2 min, a solution of 14 in DMF (1.5 mL) was added to the mixture and kept for 40 min at room temperature. The mixture was treated with Ac₂O (0.3 mL), pyridine (0.3 mL), and DMAP to give 15 (734 mg, 68%) after workup and chromatography with 4:1 hexane-EtOAc. Compound 14 had $[\alpha]_D^{23}$ -123 (c 1.02, CHCl₃); ¹H NMR data: δ 6.02-5.87 (m, 1 H, $OCH_2CH=CH_2$), 5.85 (d, 1 H, $J_{1'.2'}=4.9$ Hz, H-1'), 5.36-5.19 (m, 5 H, H-2', 3', 4', $OCH_2CH=CH_2$), 4.53 $(q, 1 \text{ H}, J_{5',6'} = 6.3 \text{ Hz}, \text{H-5'}), 4.40-4.35 \text{ (m, 1 H, OC}H_2\text{CH=CH}_2), 4.32 \text{ (d, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, OC}H_2\text{CH=CH}_2), 4.32 \text{ (d, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, OC}H_2\text{CH=CH}_2), 4.32 \text{ (d, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, OC}H_2\text{CH=CH}_2), 4.32 \text{ (d, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, OC}H_2\text{CH=CH}_2), 4.32 \text{ (d, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (d, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, OC}H_2\text{CH=CH}_2), 4.32 \text{ (d, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H, } J_{1,2} = 7.3 \text{ Hz}, \text{H-1}), 4.16-4.35 \text{ (m, 1 H,$ 4.09 (m, 1 H, OC H_2 CH=CH₂), 3.92 (dd, 1 H, $J_{5.6a}$ = 2.0 Hz, $J_{6a.6b}$ = 11.5 Hz, H-6a), 3.82 (dd, 1 H, $J_{5.6b}$ = 4.5 Hz, H-6b), 3.29-3.16 (m, 3 H, H-2, 3, 5), 2.81 (t, 1 H, $J_{3,4} = J_{4,5} = 10.4$ Hz, H-4), 2.16, 2.05, 1.98 (3 s, 9 H, 3 Ac), 1.10 (d, 3 H, H-6'), 0.94, 0.90 (2 s, 18 H, 2 t-Bu), 0.26, 0.19, 0.08 (3 s, 12 H, 4 Me); ¹³C NMR data: δ 170.5, 170.0, 169.9, 133.6, 117.5, 101.0, 79.9, 78.2, 72.9, 70.9, 69.8, 68.9, 68.4, 68.3, 65.2, 63.1, 44.4, 26.0, 25.8, 20.8, 20.6, 18.4, 18.2, 15.9, -3.1, -4.4, -5.1, -5.5; Anal. Calcd for C₃₃H₅₉N₃O₁₁SSi₂: C, 52.01; H, 7.80; N, 5.51. Found: C, 52.39; H, 7.63; N, 5.34.

Allyl 2-azido-3,6-di-O-acetyl-4-S-(2,3,4-tri-O-acetyl-α-L-fucopyranosyl)-2-deoxy-4-thio-β-D-glucopyranoside (16). A mixture of 13 (693 mg, 0,909 mmol) and M Bu₄NF solution in THF (7 mL) was stirred for 2 h at room temperature, and concentrated *in vacuo*. The residue was treated with Ac₂O (3 mL), pyridine (3 mL), and DMAP to give 16 (513 mg, 91%) after workup and chromatography with 2 : 1 hexane-EtOAc. Compound 16 had $[α]_D^{22}$ -143 (c 1.89, CHCl₃); ¹H NMR data: δ 6.00-5.86 (m, 1H, OCH₂CH=CH₂), 5.65 (d, 1 H, $J_{1',2'}$ = 5.9 Hz, H-1'), 5.37-5.23 (m, 4 H, H-2', 4', OCH₂CH=CH₂), 5.07 (dd, 1 H, $J_{2',3'}$ = 11.1 Hz, $J_{3',4'}$ = 3.1 Hz, H-3'), 4.91 (dd, 1 H, $J_{2,3}$ = 9.9 Hz, $J_{3,4}$ = 11.2 Hz, H-3), 4.54 (dd, 1 H, $J_{5,6a}$ = 2.0 Hz, $J_{6a,6b}$ = 11.9 Hz, H-6a), 4.39 (d, 1 H, $J_{1,2}$ = 8.3 Hz, H-1), 4.36-4.33 (m, 2 H, H-5', OCH₂CH=CH₂), 4.22 (dd, 1 H, $J_{5,6a}$ = 5.0 Hz, H-6b), 4.18-4.11 (m, 1 H, OCH₂CH=CH₂), 3.65 (ddd, 1 H, $J_{4,5}$ = 10.9 Hz, H-5), 3.42 (dd, 1 H, $J_{2,3}$ = 9.9 Hz, H-2), 2.92 (t, 1 H, H-4), 2.17, 2.16, 2.12, 2.06, 1.98 (5 s, 15 H, 5 Ac), 1.17 (d, 3 H, $J_{5',6'}$ = 6.6 Hz, H-6'); ¹³C NMR data: δ 170.5, 170.4, 170.0, 169.8, 169.4, 133.0, 118.3, 100.4, 82.5, 74.2, 71.1, 70.5, 70.4, 68.2, 67.2, 65.9, 65.3, 63.3, 45.4, 20.9, 20.8, 20.7, 20.6, 15.9; Anal. Calcd for C₂₅H₃₅N₃O₁₃S: C, 48.62; H, 5.71; N, 6.80. Found: C, 49.01; H, 6.23; N, 6.32.

Allyl 2-acetamido-3,6-di-O-acetyl-4-S-(2,3,4-tri-O-acetyl-α-L-fucopyranosyl)-2-deoxy-4-thio-β-D-glucopyranoside (17). To a solution of 16 (433 mg, 0.701 mmol) in aq 50% pyridine (30 mL) hydrogen sulfide was bubbled for 9 h at room temperature, then the mixture was concentrated *in vacuo*. The residue was treated with Ac₂O (5 mL), pyridine (5 mL), and DMAP to give 17 (432 mg, 97%) after workup and chromatography with EtOAc. Compound 17 had $[\alpha]_D^{24}$ -150 (c 2.09, CHCl₃); ¹H NMR data: δ 5.92-5.78 (m, 1 H, OCH₂CH=CH₂), 5.82 (d, 1 H, $J_{2,NH}$ = 9.2 Hz, NH), 5.68 (d, 1 H, $J_{1',2'}$ = 5.6 Hz, H-1'), 5.29-5.16 (m, 4 H, H-2', 4', OCH₂CH=CH₂), 5.09 (dd, 1 H, $J_{2',3'}$ = 11.0 Hz, $J_{3',4'}$ = 3.1 Hz, H-3'), 5.04 (t, 1 H, $J_{2,3}$ = $J_{3,4}$ = 10.7 Hz, H-3), 4.58 (dd, 1 H, $J_{5,6a}$ = 1.6 Hz, $J_{6a,6b}$ = 11.9 Hz, H-6a), 4.49 (d, 1 H, $J_{1,2}$ = 8.6 Hz, H-1), 4.40 (q, 1 H, $J_{5',6'}$ = 4.3 Hz, H-5'), 4.34-4.27 (m, 1 H, OCH₂CH=CH₂), 4.21 (dd, 1 H, $J_{5,6b}$ = 4.3 Hz, H-6b), 4.09 (m, 2 H, H-2, OCH₂CH=CH₂), 3.70-3.65 (m, 1 H, H-5), 3.03 (t, 1 H, $J_{4,5}$ = 10.7 Hz, H-4), 2.17, 2.12, 2.10, 2.06, 1.99, 1.95 (6 s, 18 H, 6 Ac), 1.17 (d, 3 H, H-6'); ¹³C NMR data: δ 170.7, 170.4, 170.3, 170.1, 170.0, 169.8, 133.5, 117.5, 99.6, 82.4, 74.1, 72.2, 70.5, 69.4, 68.1, 67.5, 65.7, 63.2, 55.3, 45.1, 23.1, 20.8, 20.7, 20.6, 20.5, 20.4, 15.7; Anal. Calcd for C₂₇H₃₉NO₁₄S: C, 51.18; H, 6.20; N, 2.21. Found: C, 50.95; H, 6.23; N, 2.25.

Allyl 2-acetamido-2-deoxy-4-S-(α -L-fucopyranosyl)-4-thio- β -D-glucopyrnoside (2). A solution of 17 (879 mg, 1.39 mmol) in MeOH (4 mL) and a 0.5 M solution of MeONa in MeOH (0.3 mL) were mixed, and stirred for 3 h at room temperature. The mixture was neutralized with Dowex 50W-X2 (H⁺) resin, and filtered. The filtrate was concentrate in vacuo to give 2 (381 mg, 65%). Compound 2 had mp 259-261°C (decomp.); [α]_D²⁴ -219 (c 1.08, H₂O); ¹H NMR (D₂O): δ 5.99-5.84 (m, 1 H, OCH₂CH=CH₂), 5.47 (d, 1 H, $J_{1',2'}$ = 5.6 Hz, H-1'), 5.34-5.25 (m, 2 H, OCH₂CH=CH₂), 4.55 (d, 1 H, $J_{1,2}$ = 8.6 Hz, H-1), 4.48(q, 1 H, $J_{5',6'}$ = 6.6 Hz, H-5'), 4.38-4.31 (m, 1 H, OCH₂CH=CH₂), 4.20-4.06 (m, 3 H, H-6a, 2', OCH₂CH=CH₂), 3.93 (dd, 1 H, $J_{5,6a}$ = 5.3 Hz, $J_{6a,6b}$ = 12.2 Hz, H-6b), 3.83 (d, 1 H, $J_{3',4'}$ = 2.6 Hz, H-4'), 3.76-3.57 (m, 4 H, H-2, 3, 5, 3'), 2.77 (t, 1 H, $J_{3,4}$ = $J_{4,5}$ = 10.6 Hz, H-4), 2.04 (s, 3 H, Ac), 1.21 (d, 3 H, H-6'); ¹³C NMR (D₂O): δ 174.9, 133.7, 118.5, 100.3, 85.8, 77.0, 72.0, 71.6, 70.7, 70.5, 68.1, 68.0, 61.8, 57.1, 48.0, 22.5, 15.6; Anal. Calcd for C₁₇H₂₉NO₉S: C, 48.22; H, 6.90; N, 3.31. Found: C, 48.19; H, 6.94; N, 3.28.

Allyl 2-acetamido-4,6-O-benzylidene-2-deoxy-3-O-methanesulfonyl-β-D-glucopyranoside (18). To a solution of allyl 2-acetamido-4,6-O-benzylidene-2-deoxy-β-D-glucopyranoside²⁴ (6.51 mg, 18.6

mmol) in pyridine (65 mL) was added methanesulfonyl chloride (2.9 mL, 37.5 mmol) with stirring at 0°C. After 2 h at room temperature, the mixture was poured into ice-water. The precipitate was filtered and washed with water to give 18 (2.93 g, 73%). Compound 18 had mp 200-202°C (decomp.); $[\alpha]_D^{20}$ -44 (c 0.78, CHCl₃); ¹H NMR data: δ 7.44-7.35 (m, 5 H, Ph), 5.95-5.81 (m, 2 H, OCH₂CH=CH₂, NH), 5.54 (s, 1 H, CHPh), 5.33-5.20 (m, 3 H, H-3, OCH₂CH=CH₂), 5.17 (d, 1 H, $J_{1,2}$ = 8.0 Hz, H-1), 4.39 (dd, 1 H, $J_{3,4}$ = 10.6 Hz, $J_{4,5}$ = 4.9 Hz, H-4), 4.35-4.31 (m, 1 H, OCH₂CH=CH₂), 4.15-4.08 (m, 1 H, OCH₂CH=CH₂), 3.80 (t, 1 H, $J_{5,6a}$ = 9.7 Hz, $J_{6a,6b}$ = 9.7 Hz, H-6a), 3.73 (t, 1 H, $J_{5,6b}$ = 9.7 Hz, H-6b), 3.59 (dt, 1 H, H-5), 3.47 (dt, 1 H, $J_{2,3}$ = 9.8 Hz, $J_{2,NH}$ = 8.0 Hz, H-2), 2.93 (s, 3 H, Ms), 2.03 (s, 3 H, OAc); ¹³C NMR data: δ 171.4, 136.6, 133.4, 129.4, 128.4, 126.0, 118.0, 101.8, 99.5, 79.0, 78.6, 70.7, 68.6, 65.5, 57.1, 38.5, 23.5; Anal. Calcd for $C_{10}H_{25}NO_{9}S$: C, 53.39; H, 5.89; N, 3.28. Found: C, 53.18; H, 5.98; N, 3.19.

Allyl 4,6-O-benzylidene-2,3-dideoxy-2,3-epimino-β-D-allopyranoside (19). The 3-mesylate 18 (5.92 g, 13.8 mmol) was added to a mixed solution of i-PrOH (60 mL) and dioxane (150 mL) containing metallic sodium (1.4 g, 61 mmol) dissolved under reflux. The mixture was boiled for 30 min, poured into ice water, and the product was extracted with CHCl₃. The organic layer was dried (MgSO₄) and evaporated in vacuo. The residue was chromatographed with 2:1 hexane-EtOAc to give 19 (2.93 g, 73%). Compound 19 had mp 84-85°C; [α]_D²³ -5.0 (c 1.01, CHCl₃); ¹H NMR (CDCl₃, D₂O): δ 7.52-7.30 (m, 5 H, Ph), 6.02-5.88 (m, 1 H, OCH₂CH=CH₂), 5.58 (s, 1 H, CHPh), 5.37-5.20 (m, 2 H, OCH₂CH=CH₂), 4.95 (s, 1 H, H-1), 4.38-4.30 (m, 1 H, OCH₂CH=CH₂), 4.25-4.20 (m, 2 H, H-4, OCH₂CH=CH₂), 3.79-3.64 (m, 2 H, H-6a, 6b), 2.73 (dd, 1 H, J_{2,3} = 6.6 Hz, J_{3,4} = 2.3 Hz, H-3), 2.58 (d, 1 H, H-2); ¹³C NMR data: δ 137.4, 133.8, 129.1, 128.2, 126.3, 117.6, 102.5, 98.1, 69.9, 69.3, 60.6, 34.8, 30.5; Anal. Calcd for C₁₆H₁₉NO₄: C, 66.42; H, 6.62; N, 4.84. Found: C, 66.29; H, 6.60; N, 4.89.

Allyl 4,6-O-benzylidene-2,3-dideoxy-2,3-(N-p-toluenesulfonylepimino)-β-D-allopyranoside (20). To a solution of 1 9 (2.27 g, 7.85 mmol) in pyridine (30 mL) was added TsCl (2.99 g, 15.7 mmol) with stirring at 0°C. The mixture was kept for 2 h at this temperature and the poured into ice water. The precipitate was filtered and washed with water to give 2 0 (3.40 g, 98%). Compound 20 had mp 154-155°C; $[\alpha]_D^{22}$ -12.3 (c 0.43, CHCl₃); ¹H NMR data: δ 7.87 (d, 2 H, J = 8.3 Hz, aromatic in Ts), 7.37-7.17 (m, 7 H, aromatic), 5.99-5.85 (m, 1 H, OCH₂CH=CH₂), 5.47 (s, 1 H, CH-Ph), 5.35-5.22 (m, 2 H, OCH₂CH=CH₂), 5.04 (s, 1 H, H-1), 4.36-4.29 (m, 1 H, OCH₂CH=CH₂), 4.17-4.05 (m, 2 H, H-5, OCH₂CH=CH₂), 3.96-3.92 (m, 1 H, H-4), 3.70-3.57 (m, 2 H, H-6a, 6b), 3.46 (d, 1 H, J_{2,3} =7.3 Hz, H-2), 3.27 (dd, 1 H, J_{3,4} = 2.3 Hz, H-3), 2.44 (s, 3 H, Me in Ts); ¹³C NMR data: δ 144.7, 136.9, 134.6, 133.2, 129.6, 129.1, 128.0, 128.9, 126.1, 118.1, 101.9, 96.5, 74.3, 70.3, 68.9, 61.6, 42.2, 40.3, 21.7; Anal. Calcd for C₂₃H₂₅NO₆S: C, 62.29; H, 5.68; N, 3.16. Found: C, 62.53; H, 5.79; N, 3.11.

Allyl 2,3-dideoxy-2,3-(*N*-*p*-toluenesulfonylepimino)- β -D-allopyranoside (21). A mixture of 20 (2.52 g, 1.69 mmol), Dowex 50W-X2 (H⁺) resin (9 g), and 80% MeOH (100 mL) was stirred for 2 h at 60°C. After filtration of undissolved material the filtrate was evaporated *in vacuo*. The residue was chromatographed with 1:2 hexane-BtOAc to give 21 (2.10 g, quantitative). Compound 21 had $[\alpha]_D^{22}$ -10.7 (c 3.00, CHCl₃); ¹H NMR (CDCl₃, D₂O): δ 7.84, 7.37 (each d, each 2 H, J = 8.4 Hz, aromatic in Ts), 5.94-5.79 (m, 1 H, OCH₂CH=CH₂), 5.29-5.16 (m, 2H, OCH₂CH=CH₂), 4.77 (s, 1 H, H-1), 4.33-4.25 (m, 1 H, OCH₂CH=CH₂), 4.10-4.02 (m, 1 H, OCH₂CH=CH₂), 3.98 (m, 1 H, $J_{3,4}$ = 3.5 Hz, $J_{4,5}$ = 8.6 Hz, H-4), 3.77 (m, 1H, $J_{5,6a}$ = 3.5 Hz, $J_{6a,6b}$ = 12.0 Hz, H-6a), 3.66 (m, 1H, $J_{5,6b}$ = 4.8 Hz, H-6b), 3.36 (dd, 1H, $J_{2,3}$ = 7.2 Hz, H-3), 3.31 (ddd, 1 H, H-5), 3.26 (d, 1 H, H-2), 2.46 (s, 3 H, Me in Ts); ¹³C NMR data: δ 145.2,

133.6, 133.2, 129.9, 128.1, 117.9, 95.8, 72.5, 70.0, 62.6, 62.5, 44,8, 42.8, 21.7; Anal. Calcd for $C_{16}H_{21}NO_6S$: C, 54.07; H, 5.96; N, 3.94. Found: C, 54.07; H, 6.21; N, 3.67.

 $\textbf{4,6-di-}\textit{O}\textbf{-acetyl-3-}S\textbf{-(2,3,4-tri-}\textit{O}\textbf{-acetyl-}\alpha\textbf{-L-fucopyranosyl)-2-deoxy-3-thio-}\textbf{2-p-tolu-black}$ enesulfonamido- β -D-glucopyranoside (22) and 4,6-di- θ -acetyl-2-S-(2,3,4-tri- θ -acetyl- α -Lfucopyranosyl)-3-deoxy-2-thio-3-p-toluenesulfonamido- β -D-altropyranoside (23). A mixture of 21 (604 mg, 1.69 mmol), 6 (646 mg, 1.85 mmol), and M solution of MeONa in MeOH (1.74 mL) was boiled under argon for 45 min, and then concentrated in vacuo. The residue was treated with Ac₂O (3 mL), pyridine (3 mL), DMAP to give 22 (794 mg, 63%) and 23 (362 mg, 29%) after workup and chromatographed with 3:1-1:1 hexane-EtOAc. Compound 22 had [α]_D²⁷-100 (c 3.22, CHCl₃); ¹H NMR data: δ 7.75, 7.29 (each d, each 2 H, J = 7.8 Hz, aromatic in Ts), 5.78 (d, 1 H, $J_{1',2'} = 5.7$ Hz, H-1'), 5.49-5.37 (m, 1 H, OCH₂CH=CH₂), 5.40 (d, 1 H, J_{2',3'} = 10.9 Hz), 5.26 (d, 1 H, J_{3',4'} = 3.3 Hz, H-4'), 5.11-5.02 (m, 4 H, H-3', NH, OCH₂CH=CH₂), 4.94 (t, 1 H, $J_{3,4}$ = 10.1 Hz, $J_{4,5}$ = 8.9 Hz, H-4), 4.40-4.34 (m, 2 H, H-1, H-5'), 4.20 (dd, 1 H, $J_{5,6a} = 5.1$ Hz, $J_{6a,6b} = 12.2$ Hz, H-6a), 4.07 (dd, 1 H, $J_{5,6b} = 2.6$ Hz, H-6b), 3.98-3.91 (m, 1) H, OCH₂CH=CH₂), 3.70-3.60 (m, 2 H, H-5, OCH₂CH=CH₂), 3.46 (dt, 1 H, $J_{1,2} = J_{2.NH} = 7.6$ Hz, $J_{2,3} = 1.0$ 10.3 Hz, H-2), 3.11 (t, 1 H, H-3), 2.43 (s, 3 H, Me in Ts), 2.16, 2.13, 2.09, 2.06, 1.98 (5 s, 15 H, 5 Ac), 1.15 (d, 3 H, $J_{5'6'}$ = 6.6 Hz, H-6'); ¹³C NMR data: δ 170.6, 170.4, 170.2, 169.8, 169.0, 143.3, 138.5, 133.2, 129.3, 127.2, 118.1, 100.5, 83.2, 74.1, 70.6, 69.8, 68.4, 68.3, 66.7, 65.7, 62.6, 59.6, 48.7, 21.4, 20.8, 20.7, 20.5, 15.9; Anal. Calcd for C₃₂H₄₃NO₁₅S₂: C, 51.53; H, 5.81; N, 1.88. Found: C, 51.90; H,

Compound 23 had $[\alpha]_D^{24}$ -87 (c 3.18, CHCl₃); ¹H NMR data: δ 7.75, 7.35 (each d, each 2 H, J = 8.3 Hz, aromatic in Ts), 5.92-5.80 (m, 1 H, OCH₂CH=CH₂), 5.76 (d, 1 H, $J_{1',2'}$ = 5.6 Hz, H-1'), 5.30-5.10 (m, 6 H, H-2', 3', 4', NH, OCH₂CH=CH₂), 5.01 (dd, 1 H, $J_{3,4}$ = 4.3 Hz, $J_{4,5}$ = 7.9 Hz, H-4), 4.97 (d, 1 H, $J_{1,2}$ = 2.0 Hz, H-1), 4.42 (q, 1 H, $J_{5',6'}$ = 6.6 Hz, H-5'), 4.32-4.26 (m, 1 H, OCH₂CH=CH₂), 4.19 (dd, 1 H, $J_{5,6a}$ = 5.3 Hz, $J_{6a,6b}$ = 12.1 Hz, H-6a), 4.13 (dd, 1 H, $J_{5,6b}$ = 4.1 Hz, H-6b), 4.04-3.84 (m, 3 H, H-3, 5, OCH₂CH=CH₂), 3.40 (dd, 1 H, $J_{2,3}$ = 5.6 Hz, H-2), 2.45 (s, 3 H, CH₃-C₆H₄), 2.18, 2.07, 2.06, 2.00, 1.80 (5 s, 15 H, 5 Ac), 1.17 (d, 3 H, H-6'); ¹³C NMR data: δ 170.6, 170.5, 170.1, 169.8, 168.6, 144.2, 136.4, 133.3, 129.9, 127.1, 117.7, 97.5, 84.3, 71.2, 70.9, 69.8, 68.3, 67.7, 66.2, 65.7, 62.9, 53.5, 49.3, 21.5, 20.8, 20.7, 20.6, 20.4, 15.8; Anal. Calcd for C₃₂H₄₃NO₁₅S₂: C, 51.53; H, 5.81; N, 1.88. Found: C, 51.65; H, 6.01; N, 1.75.

n-Propyl 4,6-di-O-acetyl-3-S-(2,3,4-tri-O-acetyl-α-L-fucopyranosyl)-2-deoxy-3-thio-2-p-toluenesulfonamido-β-D-glucopyranoside (24). A mixture of 2 2 (470 mg, 0.630 mmol) and 10% Pd-C (100 mg) in MeOH (5 mL) was stirred under H_2 for 24 h at room temperature, then filtered through Celite. The filtrate was concentrated in vacuo. The residue was chromatographed 1: 2 hexane-EtOAc to give 24 (486 mg, quantitative). Compound 24 had $[\alpha]_D^{24}$ -98 (c 1.90, CHCl₃); ¹H NMR data: δ 7.76, 7.30 (each d, each 2 H, J = 8.4 Hz, aromatic in Ts), 5.76 (d, 1 H, $J_{1',2'}$ = 5.6 Hz, H-1'), 5.39 (dd, 1 H, $J_{2',3'}$ = 11.0 Hz, H-2'), 5.24 (d, 1 H, $J_{3',4'}$ = 3.5 Hz, H-4'), 5.07 (dd, 1 H, H-3'), 4.99 (d, 1 H, $J_{2,NH}$ = 7.3 Hz, NH), 4.94 (t, 1 H, $J_{3,4}$ = 10.2 Hz, $J_{4.5}$ = 8.7 Hz, H-4), 4.40 (d, 1 H, $J_{1,2}$ = 7.3 Hz, H-1), 4.36 (q, 1 H, $J_{5',6'}$ = 6.3 Hz, H-5'), 4.22 (dd, 1 H, $J_{5,6a}$ = 5.1 Hz, $J_{6a,6b}$ = 12.0 Hz, H-6a), 4.07 (dd, 1 H, $J_{5,6b}$ = 3.1 Hz, H-6b), 3.62 (ddd, 1 H, H-5), 3.50 (dt, 1 H, J_d = 9.3 Hz, J_t = 7.1 Hz, OCH₂CH₂CH₃), 3.40 (dt, 1 H, $J_{2,3}$ = 10.6 Hz, H-2), 3.17 (t, 1 H, H-3), 3.00 (dt, 1 H, J_t = 6.9 Hz, OCH₂CH₂CH₃), 2.43 (s, 3 H, Me in Ts), 2.16, 2.12, 2.09, 2.06, 1.98 (5 s, 15 H, 5 OAc), 1.30-1.18 (m, 2 H, OCH₂CH₂CH₃), 1.14 (d, 3 H, H-6'), 0.70 (t, 3 H, J = 7.4 Hz, OCH₂CH₂CH₃); ¹³C NMR data: δ 170.6, 170.4, 170.2, 169.8, 169.0, 143.4, 138.6, 129.3, 127.2, 101.6,

83.1, 74.1, 71.3, 70.6, 68.5, 68.3, 66.8, 65.8, 62.7, 59.6, 48.7, 22.3, 21.4, 20.9, 20.8, 20.7, 20.5, 15.9, 10.2; Anal. Calcd for $C_{32}H_{45}NO_{15}S_2$: C, 51.40; H, 6.07; N, 1.87. Found: C, 51.27; H, 6.20; N, 1.73.

n-Propyl 2-deoxy-3-S- (α-L-fucopyranosyl)-3-thio-2-p-toluenesulfonamido-β-D-glucopyranoside (25). A solution of 24 (433 mg, 0.579 mmol) in MeOH and a 0.5 M solution of MeONa in MeOH (0.5 mL) was mixed and stirred for 20 min at room temperature. The mixture was neutralized with Dowex 50W-X2 (H⁺) resin and filtered. The filtrate was concentrated in vacuo to give 2.5 (281 mg, 90%). Compound 2.5 had mp 195-198°C; $[\alpha]_D^{24}$ -145 (c 1.06, MeOH); ¹H NMR (CD₃OD): δ 7.76, 7.32 (each d, each 2 H, J = 8.3 Hz, aromatic in Ts), 5.56 (d, 1 H, $J_{1,2}$ = 5.4 Hz, H-1'), 4.39 (q, 1 H, $J_{5,6}$ = 6.6 Hz, H-5'), 4.20 (d, 1 H, $J_{1,2}$ = 7.9 Hz, H-1), 4.07 (dd, 1 H, $J_{2',3'}$ = 9.6 Hz, H-2'), 3.85-3.30 (m, 8 H, H-2, 4, 5, 6a, 6b, 3', 4', OCH₂CH₂CH₃), 3.05-2.86 (m, 1 H, OCH₂CH₂CH₃), 2.75 (dd, 1 H, J = 9.9 Hz, J = 11.6 Hz, H-3), 1.28-1.10 (m, 2 H, OCH₂CH₂CH₃), 1.23 (d, 3 H, H-6'), 0.69 (t, 3 H, J = 7.4 Hz, OCH₂CH₂CH₃); ¹³C NMR (CD₃OD): δ 143.8, 141.6, 130.1, 128.0, 104.0, 88.2, 80.1, 73.3, 72.2, 72.1, 70.5, 69.8, 69.1, 62.9, 61.0, 55.2, 23.4, 21.4, 15.6, 10.6; Anal. Calcd for C₂₂H₃₅NO₁₀S₂: C, 49.15; H, 6.56; N, 2.61. Found: C, 48.87; H, 6.62; N, 2.40.

n-Propyl 2-acetamido-4,6-di-O-acetyl-3-S-(2,3,4-tri-O-acetyl- α -L-fucopyranosyl)-2-deoxy- β -D-glucopyranoside (26). To a solution of 25 (310 mg, 0.576 mmol) in liquid ammonia (80 mL) and THF (50 mL), cooled at -78, was added metallic sodium (300 mg). The mixture was stirred for 45 min at this temperature. After addition of NH_ACl until discharge of the blue color, liquid ammonia was distilled off, then concentrated in vacuo. The residue was treated with Ac₂O, pyridine, and DMAP to give 26 (314 mg, 86%) after workup and chromatography with 1:3 hexane-EtOAc. Compound 26 had mp 176-178°C; $[\alpha]_D^{24}$ -119 (c 1.0, CHCl₃); 5.80 (d, 1 H, $J_{2,NH}$ = 7.6 Hz, NH), 5.74 (d, 1 H, $J_{1',2'}$ = 5.5 Hz, H-1'), 5.28 (d, 1 H, $J_{3',4'}$ = 3.3 Hz, H-4'), 5.20 (dd, 1 H, $J_{2',3'} = 11.1$ Hz, H-2'), 5.09 (dd, 1 H, H-3'), 4,99 (d, 1 H, $J_{1,2} = 7.9$ Hz, H-1), 4.88 (dd, 1 H, $J_{3,4} = 11.5$ Hz, $J_{4,5} = 9.9$ Hz, H-4), 4.40 (q, 1 H, $J_{5,6} = 6.6$ Hz, H-5'), 4.23 (dd, 1 H, $J_{5,6a} = 6.6$ 5.0 Hz, $J_{6a.6b} = 11.4$ Hz, H-6a), 4.06 (dd, 1 H, $J_{5.6b} = 2.6$ Hz, H-6b), 3.81 (dt, 1 H, $J_d = 9.6$ Hz, $J_t = 6.6$ Hz, $OCH_2CH_2CH_3$), 3.76 (t, 1 H, $J_{2,3} = 11.4$ Hz, H-3), 3.74-3.68 (m, 1 H, H-5), 3.44 (dt, 1 H, $J_1 = 6.8$ Hz, OCH₂CH₂CH₃), 3.26 (dt, 1 H, H-2), 2.16, 2.08, 2.07, 2.03, 1.99 (5 s, 18 H, 6 Ac), 1.62-1.52 (m, 2 H, OCH₂CH₂CH₃), 1.18 (d, 3 H, H-6'), 0.90 (t, 3 H, J = 7.3 Hz, OCH₂CH₂CH₂CH₃); ¹³C NMR data: δ 170.8, 170.7, 170.4, 170.3, 169.8, 169.2, 100.1, 82.0, 74.2, 71.4, 70.5, 68.3, 68.2, 67.7, 65.2, 62.6, 58.6, 45.9, 23.5, 22.6, 20.8, 20.7, 20.6, 20.5, 16.0, 10.3; Anal. Calcd for $C_{22}H_{43}NO_{14}S$: C, 51.02; H, 6.50; N, 2.20. Found: C, 50.97; H, 6.52; N, 2.20.

n-Propyl 2-acetamido-2-deoxy-3-S-(α-L-fucopyranosyl)-β-D-glucopyranoside (3). A solution of 26 (326 mg, 0.513 mmol) in MeOH (1 mL) and a 0.5 M solution of MeONa in MeOH (1 mL) were mixed and stirred for 1 h at room temperature was concentrated in vacuo to give 27 (206 mg, 94%). Compound 3 had mp 247-248°C; $[\alpha]_D^{24}$ -220 (c 0.45, H₂O); ¹H NMR (D₂O): δ 5.38 (d, 1 H, $J_{1',2'}$ = 5.6 Hz, H-1), 4.53 (d, 1 H, $J_{1,2}$ = 8.3 Hz, H-1), 4.68 (q, 1 H, $J_{5',6'}$ = 6.3 Hz, H-5'), 4.04 (dd, 1 H, $J_{2',3'}$ = 10.6 Hz, H-2'), 3.94 (dd, 1 H, $J_{5,6a}$ = 1.7 Hz, $J_{6a,6b}$ = 12.3 Hz, H-6a), 3.92-3.81 (m, 3 H, H-2, 4', OCH₂CH₂CH₃), 3.76 (dd, 1 H, $J_{5,6b}$ = 4.6 Hz, H-6b), 3.71 (dd, 1 H, $J_{3',4'}$ = 3.5 Hz, H-3'), 3.61-3.51 (m, 2 H, H-5, OCH₂CH₂CH₃), 3.45 (t, 1 H, $J_{3,4}$ = $J_{4,5}$ = 9.6 Hz, H-4), 2.80 (t, 1 H, H-3), 2.05 (s, 3 H, Ac), 1.60-1.52 (m, 2 H, OCH₂CH₂CH₃), 1.21 (d, 3 H, H-6'), 0.87 (t, 3 H, $J_{5,5}$ = 7.4 Hz, OCH₂CH₂CH₃); ¹³C NMR (D₂O): δ 174.5, 102.5, 87.5, 78.7, 72.5, 72.0, 70.5, 68.1, 67.8, 61.5, 55.8, 52.8, 22.4, 15.7, 10.0; Anal. Calcd for C₁₇H₃₁NO₉S: C, 47.99; H, 7.34; N, 3.29. Found: C, 47.51; H, 7.45; N, 3.08.

3, 4-Di-O-acetyl-2-S-(2, 3, 4-tri-O-acetyl- α -L-fucopyranosyl)-1, 6-anhydro-2-thio- β -Dgalactopyranose (28) and 2,4-di-O-acetyl-3-S-(2,3,4-tri-O-acetyl-\alpha-L-fucopyranosyl)-1,6anhydro-3-thio-β-D-idopyranose (29). A mixture of 6 (383 mg, 1.10 mmol), 2 7²⁵ (144 mg, 1.00 mmol), and a 2.9 M solution of MeONa in MeOH (0.38 mL) was stirred for 2 h at 95°C under argon. The mixture was treated with Ac₂O (2 mL), pyridine (2 mL), and DMAP to give 28 (465 mg, 87%) and 29 (39 mg, 7%) after workup and chromatography with 2:1-1:1 hexane-EtOAc. Compound 28 had mp 173-174 °C; $[\alpha]_D^{24}$ -187 (c 1.46, CHCl₃); ¹H NMR data: δ 5.95 (d, 1 H, $J_{1',2'}$ = 5.3 Hz, H-1'), 5.48 (s, 1 H, H-1), 5.34 (t, 1 H, $J_{3,4} = 5.0$ Hz, $J_{4,5} = 4.3$ Hz, H-4), 5.31 (dd, 1 H, $J_{2',3'} = 11.0$ Hz, H-2'), 5.29 (d, 1 H, $J_{3',4'} = 3.3$ Hz, H-4'), 5.21 (dd, 1 H, $J_{2.3}$ = 1.0 Hz, H-3), 5.14 (dd, 1 H, H-3'), 4.49-4.45 (m, 2 H, H-5, 5'), 4.41 (d, 1 H, $J_{6a,6b}$ = 7.3 Hz, H-6a), 3.74 (dd, 1 H, $J_{5,6d}$ = 5.6 Hz, H-6b), 3.13 (s, 1 H, H-2), 2.16, 2.11, 2.10, 2.06, 1.99 (5 s, 15 H, 5 Ac), 1.16 (d, 3 H, $J_{5'.6'} = 6.6$ Hz, H-6'); ¹³C NMR data: δ 170.4, 170.0, 169.9, 169.8, 169.2, 101.9, 82.4, 72.5, 70.8, 70.7, 68.4, 67.4, 65.5, 64.9, 64.8, 46.9, 20.9, 20.7, 20.6, 20.5, 15.9; Anal. Calcd for C₂₂H₃₀O₁₃S: C, 49.43; H, 5.66. Found: C, 49.32; H, 5.66. Compound 29 had mp 159-160°C; $[\alpha]_D^{24}$ -203 (c 0.76, CHCl₃); ¹H NMR data: δ 5.93 (d, 1 H, $J_{1',2'} = 5.0$ Hz, H-1'), 5.45 (d, 1 H, $J_{1,2} = 1.5$ Hz, H-1), 5.30 (d, 1 H, $J_{3',4'} = 2.0$ Hz, H-4'), 5.20-5.13 (m, 2 H, H-1) 2',3'), 5.08 (dd, 1 H, $J_{3,4}$ = 10.1 Hz, $J_{4,5}$ = 3.5 Hz, H-4), 4.67 (dd, 1 H, $J_{2,3}$ = 10.5 Hz, H-2), 4.56 (t, 1 H, $J_{5.6b} = 5.0 \text{ Hz}$, H-5), 4.47 (q, $J_{5'.6'} = 6.6 \text{ Hz}$, H-5'), 4.10 (d, 1 H, $J_{6a,6b} = 7.9 \text{ Hz}$, H-6a), 3.76 (dd, 1 H, H-6a) 6b), 3.19 (t, 1 H, H-3), 2.17, 2.13, 2.04, 2.00 (4 s, 15 H, 5 Ac), 1.21 (d, 3 H, H-6'); ¹³C NMR data: δ 170.4, 170.3, 169.9, 169.8, 169.7, 99.1, 82.4, 73.9, 73.0, 72.9, 70.6, 68.2, 68.0, 65.4, 64.9, 41.4, 20.8,

1,3,4,6-Tetra-O-acetyl-2-S-(2,3,4-tri-O-acetyl- α -L-fuc opyranosyl)-2-thio-D-galactop yranose (30). A solution of 28 (1.26 g, 2.36 mmol) in Ac₂O-AcOH-H₂SO₄ (7:3:0.07 v/v; 3 mL) was stirred at room temperature for 1 h. Sodium acetate (500 mg) was then added and the solution was concentrated *in vacuo*. The residue was dissolved in EtOAc and the solution was washed with brine dried (MgSO4), and evaporated in vacuo. The residue was chromatographed with 1:1 hexane-EtOAc to give 30 (1.56 g, quantitative) as a 88:12 mixture of α and β anomers. Compound 30 had mp 219-221°C; $[\alpha]_D^{22}$ -119 (c 1.13, CHCl₃); ¹H NMR data: δ 6.21 (d, 1 H, $J_{1,2}$ = 3.6 Hz, H-1), 5.88 (d, 1 H, $J_{1',2'}$ = 4.6 Hz, H-1'), 5.39 (d, 1 H, $J_{3,4}$ = 3.6 Hz, H-4), 5.28-5.27 (m, 1 H, H-4'), 5.27 (dd, 1 H, $J_{2,3}$ = 11.5 Hz, H-3), 5.19-5.09 (m, 2 H, H-2', 3'), 4.43 (q, 1 H, $J_{5',6'}$ = 6.6 Hz, H-5'), 4.27 (t, 1 H, $J_{5,6}$ = 6.9 Hz, H-5), 4.08(d, 2 H, H-6), 3.35(dd, 1 H, H-2), 2.19, 2.16, 2.15, 2.03, 2.02, 1.99 (6 s, 21 H, 7 Ac), 1.20 (d, 3 H, H-6'); ¹³C NMR data: δ 170.4, 170.3, 170.1, 170.0, 169.8, 168.8, 92.7, 83.4, 70.7, 70.3, 68.6, 68.1, 68.0, 66.7, 65.1, 61.2, 42.0, 20.8, 20.6, 20.5, 20.4, 15.9; Anal. Calcd for C₂₆H₃₆O₁₆S: C, 49.05; H, 5.70. Found: C, 48.86; H, 5.81.

20.7, 20.6, 20.5, 15.8; Anal. Calcd for C₂₂H₃₀O₁₃S: C, 49.43; H, 5.66. Found: C, 49.15; H, 5.44.

 $O-{3,4,6-Tetra-O-acetyl-2-S-(2,3,4-tri-O-acetyl-α-L-fucopyranosyl)-2-thio-α-D-galactopyranosyl}$ trichloroacetimidate (31). A mixture of 30 (617 mg, 0.812 mmol) and NH₂NH₂-AcOH (152 mg, 1.65 mmol) in DMF was stirred for 20 min at 50°C, and diluted with EtOAc. The mixture was washed with brine, dried (MgSO₄), and evaporated *in vacuo*. The residue was chromatographed with 3:5 hexane-EtOAc to give the corresponding hemiacetal (372 mg, 65%). A solution of this hemiacetal (543 mg, 0.913 mmol) in CH₂Cl₂ (1 mL), stirred under argon, was treated successively with CCl₃CN (0.90 mL, 9.0 mmol) and DBU (50 μL) at 0°C. After being stirred for 1 h at 0°C, the mixture was directly chromatographed with 4:1-1:1 hexane -EtOAc to give 31 (598 mg, 89%). Compound 31 had mp 181-182 °C; [α]_D-84 (c 1.07, CHCl₃); ¹H NMR data: δ 8.78 (s, 1 H, NH), 6.41 (d, 1 H, $J_{1,2}$ = 3.3 Hz, H-1), 5.91 (d,

1 H, $J_{1',2'}$ = 4.5 Hz, H-1'), 5.45 (d, 1 H, $J_{3,4}$ = 3.1 Hz, H-4), 5.33 (dd, 1 H, $J_{2,3}$ = 11.7 Hz, H-3), 5.31 (m, 1 H, H-4), 5.19 (dd, 1 H, $J_{2',3'}$ = 10.8 Hz, H-2'), 5.12 (dd, 1 H, $J_{3,4}$ = 2.8 Hz, H-3'), 4.49 (q, 1 H, $J_{5',6'}$ = 6.3 Hz, H-5'), 4.42 (t, 1 H, $J_{5,6a}$ = 6.3 Hz, $J_{5,6b}$ = 7.3 Hz, H-5), 4.14 (dd, 1 H, $J_{6a,6b}$ = 11.5 Hz, H-6a), 4.07 (dd, 1 H, H-6b), 3.43 (dd, 1 H, H-2), 2.17, 2.16, 2.04, 2.03, 2.01, 1.99 (6 s, 18 H, 6 Ac), 1.21 (d, 3 H, H-6'); 13 C NMR data: 8 170.4, 170.3, 170.2, 170.0, 169.9, 169.8, 160.6, 96.8, 83.4, 70.6, 70.4, 69.2, 68.2, 68.1, 66.8, 65.0, 61.4, 42.4, 20.7, 20.6, 20.5, 20.4, 15.9; Anal. Calcd for $C_{26}H_{34}C_{13}NO_{15}S$: C, 42.26; H, 4.64; N, 1.90. Found: C, 42.25; H, 4.76; N, 1.80.

3,4,6-tri-O-acetyl-2-S-(2,3,4-tri-O-acetyl-α-L-fucopyranosyl)-2-thio-β-D-galactopy-Allyl To a mixture of 31 (816 mg, 1.10 mmol), allyl alcohol (0.45 mL, 6.6 ranoside (32) and α -anomer. mmol), and MS 4A (2 g), stirred under argon at -45°C, was added a solution of BF₃·OEt₂ (34 mL, 0.28 mmol) in CH₂Cl₂ (0.3 mL) and filtered through Celite. The filtrate was evaporated in vacuo. The residue was chromatographed with 1:1-1:2 hexane-EtOAc to give 32 (634 mg, 90%) and its α -anomer (56 mg, 8%). Compound 33a had $[\alpha]_D^{21}$ -189 (c 1.15, CHCl₂); ¹H NMR data: δ 6.03-5.92 (m, 1 H, OCH₂CH=CH₂), 5.90 (d, 1 H, $J_{1',2'}$ = 5.1 Hz, H-1'), 5.38-5.19 (m, 5 H, H-4, 3', 4', OCH₂CH=C H_2), 5.14 (dd, 1 H, $J_{2',3'}$ = 10.7 Hz, H-2'), 4.99 (dd, 1 H, $J_{2,3} = 11.8$ Hz, $J_{3,4} = 3.3$ Hz, H-3), 4.60 (q, 1 H, $J_{5',6'} = 6.6$ Hz, H-5'), 4.41 (d, 1 H, $J_{1.2} = 9.1$ Hz, H-1), 4.41-4.36 (m, 1 H, OC H_2 CH=CH₂), 4.19 (dd, 1 H, $J_{5.6a} = 6.6$ Hz, $J_{6a,6b} = 11.0$ Hz, H-6a), 4.14-4.11 (m, 1 H, OC H_2 CH=CH₂), 4.12 (dd, 1 H, $J_{5.6b}$ = 6.9 Hz, H-6b), 3.85 (t, 1 H, H-5), 3.09 (dd, 1 H, H-2), 2.16, 2.13, 2.05, 2.03, 2.01, 2.00 (6 s, 18 H, 6 Ac), 1.16 (H-6'); ¹³C NMR data: δ 170.4, 170.3, 170.2, 170.1, 170.0, 169.9, 133.3, 118.6, 100.5, 82.4, 73.7, 70.9, 70.8, 70.4, 68.4, 68.2, 66.1, 65.1, 61.4, 44.1, 20.6, 20.5, 20.3, 15.7; Anal. Calcd for C₂₇H₃₈O₁₅S: C, 51.10; H, 6.04. Found: C, 51.20; H, 6.40.

Compound 33b had mp 162-163°C; $[\alpha]_D^{21}$ -107 (c 0.58, CHCl₃); 5.99 (m, 2 H, H-1', OCH₂CH=CH₂), 5.38-5.17 (m, 7 H, H-3, 4, 2', 3', 4', OCH₂CH=CH₂), 4.93 (d, 1 H, $J_{1,2}$ = 3.4 Hz, H-1), 4.47 (q, 1 H, $J_{5',6'}$ = 6.6 Hz, H-5'), 4.26-4.00 (m, 5 H, H-5, 6a, 6b, OCH₂CH=CH₂), 3.30 (dd, 1 H, $J_{2,3}$ = 11.2 Hz, H-2), 2.16, 2.12, 2.05, 2.02, 2.01, 2.00 (6 s, 18 H, 6 Ac), 1.19 (d, 3 H, $J_{5',6'}$ = 6.6 Hz, H-6'); ¹³C NMR data: δ 170.4, 170.2, 170.1, 169.9, 133.0, 118.3, 71.5, 70.7, 69.0, 68.1, 67.4, 66.8, 64.7, 61.9, 42.0, 20.6, 20.5, 20.4, 15.9, Anal. Calcd for $C_{27}H_{38}O_{15}S$: C, 51.10; H, 6.04. Found: C, 51.00; H, 6.04.

Allyl 2-S-(α-L-fucopyranosyl)-2-thio-β-D-galactopyranoside (4). A solution of 32 (596 mg, 0.939 mmol) in MeOH (2 mL) and a 0.5 M solution of MeONa in MeOH (0.3 mL) were mixed and stirred for 2 h at room temperature. The mixture was neutralized with Dowex 50W-X2 (H⁺) resin and filtered. The filtrate was concentrated *in vacuo* to give 4 (371 mg, quantitative). Compound 34 had mp 171-172°C; $[\alpha]_D^{21}$ -177 (c 0.66, H₂O); ¹H NMR data: δ 6.08-5.98 (m, 1 H, OCH₂CH=CH₂), 5.59 (d, 1 H, $J_{1',2'}$ = 5.5 Hz, H-1'), 5.43-5.27 (m, 2 H, OCH₂CH=CH₂), 4.56 (d, 1 H, $J_{1,2}$ = 8.9 Hz, H-1), 4.46 (q, $J_{5',6'}$ = 5.6 Hz, H-5'), 4.39-4.37 (m, 1 H, OCH₂CH=CH₂), 4.30-4.23 (m, 1 H, OCH₂CH=CH₂), 4.08 (dd, 1 H, $J_{2',3'}$ = 10.2 Hz, H-2'), 3.90-3.66 (m, 7 H, H-3, 4, 5, 6a, 6b, 3', 4'), 2.83 (t, 1 H, $J_{2,3}$ = 10.9 Hz, H-2), 1.22 (d, 3 H, H-6'); ¹³C NMR data: δ 134.0, 118.9 (OCH₂CH=CH₂), 101.4, 86.9, 75.3, 73.8, 72.1, 71.1, 70.7, 68.4, 68.2, 68.0, 61.4, 49.0, 15.9 Anal. Calcd for C₁₅H₂₆O₉S: C, 47.11; H, 6.85. Found: C, 47.11; H, 7.14.

Measurement of inhibition activity. α -L-Fucosidases from bovine kidney and bovine epididymis were purchased from Sigma Chemical Co. and p-nitrophenyl α -L-fucopyranoside from Seikagaku Kogyo Co. Enzyme assay was performed by essentially the same method as that of Evans *et al.* ²⁶ Inhibition assay was performed at 25°C for 30-40 min in 20 mM citrate buffer (pH 5.5 for bovine kidney and pH 5.8 for bovine

epididymis, 300 μ L), which contained the following assay components: p-nitrophenyl α -L-fucopyranoside (0.07-0.33 mM), the S-linked disaccharide 1, 2, 3 (each 0-6.7 mM, two intermediate points) or 4 (0-1 mM, two intermediate points), α -L-fucosidase (0.8-1.1 units/mL for bovine kidney and 1.4 units/mL for bovine epididymis). After addition of 50 mL glycine buffer (pH 10.0, 500 μ L), UV absorption at 400 nm was measured. The data expressed by Lineweaver-Burk plot and replots of the slope versus the inhibitor concentration and the intercept versus the inhibitor concentration indicate the modes of inhibition and give the Ki values as well as the coefficient α .

REFERENCES AND NOTES

- † Present address: Department of Biological Engineering, Science University of Tokyo, 2641 Yamazaki, Nodashi, 278 Japan
- Defaye, J.; Gelas, J. in Studies in Natural Product Chemistry; Atta-ur-Rahman Ed; Vol. 8, Elsevier, Amsterdam, 1991, pp. 315-357.
- 2. Blanc-Muesser, M.; Vigne, L.; Driguez, H.; Lehmann, J.; Steck, J.; Urbahns, K. Carbohydr. Res., 1992, 224, 59-71.
- (a) Kobata, A. The Carbohydrates of Glycoproteins. In Biology of Carbohydrates; Ginsburg, V.; Robins P.
 W. Eds.; John Wiley and Sons, Inc., New York, 1984; Vol. 2, pp. 87-161; (b) Feizi, T. Nature, 1985, 314, 53-57; (c) Hakomori, S.; Adv. Cancer Res., 1989, 52, 257-331.
- (a) DeSantis, R.; Pinto, M. R. Gamete Interaction in Ascidians: Sperm Binding and Penetration Through the Vitelline Coat. in *Mechanism of Fertilization: Plants to Humans*, Dale, B. Ed.; Springer-Verlag: Berlin, 1990, pp. 297-304; (b) Hoshi, M.; DeSantis, R.; Pinto, M. R.; Cotelli, F.; Rosati, F. Zool. Sci., 1985, 2, 65-69.
- 5. Huang Jr., T. F.; Ohzu, E.; Yanagimachi, R. Gamete Res., 1982, 5, 355-361.
- 6. Dicioccio, R. A.; Barlow, J. J.; Matta, K. L. J. Biol. Chem., 1982, 257, 714-718.
- (a) Paulsen, H. Angew. Chem., Int. Ed. Engl., 1981, 21, 155-224; (b) Schmidt, R. R. Angew. Chem., Int. Ed. Engl., 1986, 25, 212-235; (c) A recent review including many other reviews: Toshima, K.; Tatsuta, K. Chem. Rev., 1993, 93, 1503-1531.
- 8. Matta, K. L.; Girotra R. N.; Barlow, J. J. Carbohydr. Res., 1975, 43, 101-109.
- (a) Farkas, I; Szabó I. F.; Bognár, R. Carbohydr. Res., 1976, 48, 136-138; (b) Blanc-Muesser, M.;
 Defaye, J.; Driguez, H. J. Chem. Soc. Perkin Trans. I, 1982, 15-18.
- 10. Gadelle, A.; Defaye, J.; Pederson, G.; Carbohydr. Res., 1990, 200, 497-498.
- 11. Unpublished result by Hashimoto, H.; Kanazawa, M.; Horito, S.: Nucleophilic substitution of methyl 2-acetamido-2-deoxy-3,6-di-O-pivaloyl-4-O-triflyl-β-D-galactopyranoside with DMF at room temperature for 1 h gave the desired 4-acetylthio derivative in 50% yield. A longer reaction time or elevated temperature promoted the elimination reaction to give the corresponding 4-enopyranoside.
- 12. Yamaguchi, T. Carbohydr. Res., 1983, 119, 279-284.
- 13. Mayer zu Reckendorf, W.; Lenzen, H.-J. Liebigs Ann. Chem., 1985, 477-484.
- (a) Tipson, R. S. in Methods in Carbohydrate Chemistry; Whistler, R. L.; Wolfrom, M. L. Eds; Vol. 2, Academic Press, New York, 193, pp 250-251; (b) Jarell, H. C.; Ritchie, R. G. S.; Starek, W. A.; Jones, J. K. N. Can. J. Chem., 1973, 51, 1767-1770; (c) Binkley, R. W.; Koholic, D. J. J. Org. Chem., 1989,

- 54, 3577-3581; (d) Gold, E. H.; Badad, E. J. Org. Chem., 1972, 37, 2208-2210.
- 15. Knapp, C.; Lehmann, J.; Schwesinger, B. Chem. Ber., 1979, 112, 1147-1152.
- 16. Williams, N. R. Adv. Carbohydr. Chem. Biochem., 1970, 25, 109-179.
- 17. Hardegger, E.; Schuep, W. Helv. Chim. Acta, 1970, 53, 951-959.
- Recent publications: (a) Schudder, P.; Naville, D. C. A.; Butters, T. D.; Fleet, G. W. J.; Dwek, R. A.; Rademacher, T. W.; Jacob, G. S. J. Biol. Chem., 1990, 265, 16472-16477; (b) Svensson, S. C. T.; Thiem, J. Carbohydr. Res., 1990, 200, 391-402; (c) Tsuji, Y.; Yamamoto, K.; Tochikura, T. J. Biochem., 1990, 108, 235-240; (d) Tsuji, Y.; Yamamoto, K.; Tochikura, T.; Seno, T., Ohkubo, Y.; Yamaguchi, H.; J. Biochem., 1990, 107, 324-330; (e) Sano, M.; Hayakawa, K.; Kato, I. J. Biol Chem., 1992, 267, 1522-1527.
- (a) Srivastava, P. N.; Arban, K.; Takei, G. H.; Huang, T. T. F.; Yamaguchi, R. Biochem. Biophys. Res. Commun., 1986, 137, 1061-1068; (b) Jauhiainen, A; Vanha-Perttula, T. Biochem Biophys Acia, 1986, 880, 91-95.
- 20. Hashimoto, H.; Izumi, M. Tetrahedron Lett., 1993, 31, 4949-4952.
- 21. Segel, I. H. in *Enzyme Kinetics*, John Wiley & Sons, New, York, 1975, pp. 161-202.
 The coefficient α is defined as shown in the following system of the linear mixed-type inhibition, where E,
 S, I, and P mean enzyme, substrate, inhibitor, and product, respectively.

in enzyme, substrate, inhibitor, and product, respectively.

$$E + S \xrightarrow{K_s} ES \xrightarrow{k_p} E + P$$

$$+ + K_s = [E][S] / [ES]$$

$$K_i \downarrow \downarrow \qquad \qquad K_i = [E][I] / [EI]$$

$$K_i \downarrow \downarrow \qquad \qquad \alpha K_i \downarrow \downarrow \qquad \qquad \nu = k_p[ES]$$

$$EI + S \xrightarrow{\alpha K_s} ESI$$

- 22. Lee, B. T.; Lee, Y. C. Carbohydr. Res., 1974, 37, 193-201.
- 23. Lemieux, R. V.; Ratcliffe, R. M. Can. J. Chem., 1979, 57, 1244-1251.
- 24. Shaban, M. A. E.; Reinhold, V. N.; Jeanloz, R. W. Carbohydr. Res., 1977, 59, 213-233.
- 25. Jeanloz, R. W.; Stoffyn, P. in *Methods in Carbohydrate Chemistry*; Whistler, R. L.; Wolfrom, M. L. Eds.; Vol. 1, Academic Press, New York, 1962, pp. 221-227.
- 26. Evans, S.V.; Fellows, L. E.; Shing, T. K. M.; Fleet, G. W. J. Phytochemistry, 1985, 24, 1953-1955.

(Received 20 September 1994)